

British Microbiology Research Journal 5(1): 52-56, 2015, Article no.BMRJ.2015.006 ISSN: 2231-0886



SCIENCEDOMAIN international www.sciencedomain.org

Carrier Rate of Typhoid Organisms in Enugu, Nigeria

Martin E. Ohanu^{1*} and Bryan O. Ogeneh¹

¹Department of Medical Microbiology, College of Medicine, University of Nigeria, Enugu Campus, Nigeria.

Authors' contributions

This work was carried out in collaboration between both authors. Both authors read and approved the final manuscript.

Article Information

DOI: 10.9734/BMRJ/2015/12421 <u>Editor(s):</u> (1) Gadi Borkow, Chief Medical Scientist Cupron Scientific, Israel. (1) Anonymous, University of Agriculture, Nigeria. (2) Anonymous, Umm Al-Qura University, Saudi Arabia. Peer review History: <u>http://www.sciencedomain.org/review-history.php?iid=657&id=8&aid=6039</u>

Short Communication

Received 30th June 2014 Accepted 21st August 2014 Published 10th September 2014

ABSTRACT

Aims: To determine the carrier rate of typhoid and paratyphoid organisms in Enugu community. **Place and Duration of Study:** The project was carried out in Enugu Urban in South Eastern part of Nigeria. Those enrolled in the study that lasted for one year are students, traders and civil servants.

Methodology: Ninety six apparently healthy adults without any complaints, comprising of forty eight males and forty eight females were selected for the study. Blood, urine and stool samples were collected from all in the study group. The samples were subjected to standard bacteriological culture for enteric organisms at the Medical Microbiology Laboratory of University of Nigeria Teaching Hospital, Enugu.

Results: Salmonella typhi was isolated from the stool of two (2.08%) of the candidates while *S. paratyphi* was recovered from the stool of another individual (1.04%) of the study group. No candidate gave Widal titre greater than 40 to 'O' agglutinin which is statistically significant (Anova F value 597.7, P value <0.0001). Only Two Candidates positive for *S. typhi* culture gave titre of 160 to 'H' antigen, also Statistically significant (Anova F value 1195, P value <0.0001).

Conclusion: Typhoid carrier rate in Enugu Community is 2.1% while that of paratyphoid is 1.0%. From the study females are more likely to be typhoid carriers in Enugu.

Keywords: Typhoid carrier; bacteriology; Widal test.

1. INTRODUCTION

Infection with Salmonella organisms remain a world-wide problem. Salmonella typhi is a serious source of illness primarily in the developing world, with over 12.5 million cases estimated outside China in 1980 [1]. Salmonellosis is generally acquired through contaminated food and water, although cross contamination occur in hospitals and institutions [2]. Nearly 50% of determined outbreaks are caused by poultry and poultry products. Typhoid fever remains an important public health problem with more than half million deaths occurring throughout the developing world [3]. In Nigeria, up to 30 percent of the patients admitted may die from the infection [3]. A factor of epidemiologic importance is the asymptomatic human carrier state (chronic carrier and temporary excreter), which exist for the agents of typhoidal and nontyphoidal salmonellosis. [4] Approximately, 3 % of persons infected with S. typhi and 0.1% of those infected with non-typhoidal salmonellae become chronic carriers which can last from many weeks to years. In chronic typhoid carriers, persistent carriage of very large numbers of organisms is usual- 106-108 organisms/g of faeces. [5] Children rarely become chronic typhoid carriers⁴. The chronic carrier state is defined as documented excretion of S. typhi in stool or urine for a year or more [5]. About one third of the chronic carriers give no history consistent with typhoid fever and the incidence is higher in older patients and more in women. [6] Underlying biliary or urinary tract diseases, especially with stone formation, increase the probability of the chronic enteric or urinary carrier states in patients with typhoid fever [5]. In Nigeria typhoid fever patients are diagnosed on the basis of clinical symptoms and a single Widal test [7]. Bacteriologic culture is seldom done. Vi (capsuler antigens of S. typhi) agglutinins are found in the serum of some 85% of chronic carriers at a titre of 5 or more. [8] The measurement of antibodies to Vi by haemagglutination test and by EIA (enzyme immunosobent assay) commonly called Enzyme Linked Immunosorbent Assay (ELISA) can detect chronic S. typhi carriers if bacteriologic culture cannot be done [8]. Vaccines against typhoid are available the parenteral vi capsular polysaccharide and the oral live whole-cell Salmonella typhi 21a vaccine that provide similar levels of protection. [9] The response to Ty21a vaccine was found to imitate that in natural infection. [10] Worldwide, nontyphoid salmonella (NTS) create a huge public health concern with155,000 deaths and over 90 million cases annually.¹¹ They are known for their emerging antimicrobial resistance and there are no vaccines against them. The 21a vaccine elicited a higher immune response cross-reactive with several NTS strains and is at the forefront as a surrogate vaccine [11]. There is little report of typhoid carrier rate in Enugu Nigeria. This study aims to determine the carrier rate in this community.

2. MATERIALS AND METHODS

Ninety-six apparently healthy adults (48 males and 48 females) without any complaints during the period of study, were selected for the study. In the group were traders, students and civil servants in Enugu. Blood, urine and stool samples were collected from the candidates. Stool and urine specimens were inoculated onto appropriate bacteriologic agars (MacConkey and Deoxycholate agars) before and after enrichment in selenit-F broth (1gm or 1ml of stool) that was incubated for 24 hrs. Subcultures of stool and urine were plated onto MacConkey and Deoxycholate agars. Urine samples were additionally inoculated on Cystein Lactose Electrolyte Deficient (CLED) agar. The resulting Non Lactose fermenting suspect colonies on the above agars that did not metabolize urea but produced hydrogen sulphide in Bismuth sulphite medium were examined for salmonella including serologic typing by standard methods [12]. Widal agglutination tests were done on the sera resulting from the blood samples. The procedure used was the standard tube dilution method of described by Ismail [13] with Freter as salmonella antigens prepared by Gama Biological.

3. RESULTS

All the 96 candidates were screened for typhoid and other enteric fever organisms. *Salmonella typhi* was isolated from the stool of two (2.08%) of the candidates, while *S. paratyphi C* was recovered from the stool of another individual (1.04%) of the study group (Table 1). With regards to Widal test, none of them gave a titre greater than 40 for '0' agglutinin which is statistically significant (Anova test F value 597.7, P value <0.0001). The two from whom *S. typhi* was recovered, gave titre of 160 against 'H' antigen also statistically significant (Anova F value 1195, P value <0.0001) see (Table 2). The two typhoid isolates came from females while the paratyphi organism came from a male. Repeat stool cultures from the three culture positive individuals gave similar results after a period of six and twelve months, confirming that they are carriers.

Table 1. Isolation rate, age range and sex distribution of the individuals culture positive for enteric fever organisms (%)

Sex	S. typhi	S. paratyhi C	Age
Male	0(0)	1 (1.04)	27
Female	2(2.08)	O (0)	30 – 32

4. DISCUSSION

From the results typhoid organisms were recovered from three apparently healthy individuals, even on repeat cultures. The study group were considered healthy since they gave no complaints. The rate of 2.1 percent should be considered the typhoid carrier rate in Enugu. Also 1.0 percent should be taken as the paratyphi salmonella carrier rate.

Typhoid, an illness caused by *Salmonella enteric serovar typhi* is common worldwide; it is transmitted by the ingestion of food or water contaminated with faeces from an infected person [4]. When untreated typhoid fever persists for three weeks to a month [14]. Azithromycin and fluoroquinolone are antibiotics suggested for treatment [14]. Death occurs in between 10% and 47% of untreated cases [15]. A person may become an asymptomatic carrier of typhoid fever, suffering no symptoms, but capable of infecting others. Approximately 5% of people who contract typhoid continue to carry the causative organism after they recover (like Mary Mallon-typhod Mary) [16]. With a global estimate of 16-33 million cases annually resulting in half a million deaths, typhoid is identified as a serious public health problem [15]. Some patients come to the clinic with typhoid "certificates" as evidenced by their previous positive laboratory Widal results [7]. This can contribute to over diagnosis of typhoid [7]. Due to improved surgery and supportive care in some countries, median case fatality rate (CFR) for typhoid perforation (TP) declined from over 50% in 1960 to single digits in the last decade but West Africa lags behind at 16% [17]. There is mounting evidence of the association between hepatobilliary cancer and typhoid carrier state [18,19]. Though some regard Kathmandu, Nepal as the enteric fever capital of the world [20], the global burden of typhoid fever is not in doubt [21]. Public health education campaigns encouraging people to wash their hands after defecating and before handling food are an important component in controlling spread of the disease. Control of carriers can be further done through encouraging them to submit to chemotherapy or be considered for cholecystectomy [19]. It is recommended that health authorities should keep a register of typhoid and paratyphoid carriers. The carriers themselves should be kept under "surveillance" by local health officers who should ensure that the sewage from their dwellings is safely disposed of, give them appropriate instructions in personal hygiene and encourage them not to take employment in food or water industry. However workers in the food or water industry should be individually examined bacteriologically or culture of sewage effluent of large food factories can be done as an alternative to the examination of individual faecal specimen [21]. Chlorination of drinking water, adequate treatment and vaccination against the disease has led to dramatic decreases in the transmission of typhoid fever in the U.S [22].

Table 2. Widal test for (96) healthy normal individual
--

Antigens					Titre			
-		< 20	20	40	80	160	320	640
'O'	{DO (S. typhi)	96	0	0	0	0	0	0
Antigen	{aO S. paratyphi A	96	0	0	0	0	0	0
	{bO S. paratyphi B	86	2	8	0	0	0	0
	{cO S. paratyphi C	86	3	7	0	0	0	0
'H'	{DH	87	7	0	0	2	0	0
Antigen	{aH	96	0	0	0	0	0	0
	{bH	95	1	0	0	0	0	0
	{cH	95	1	0	0	0	0	0

Both paratyphi A and B have no vaccine in clinical use though contentiously Ty 21a-vaccine have been suggested and there is a growing problem of antibiotic resistance among the strains [23]. Strains of *Salmonella enteric* sub sp enteric are amongst the most commonly identified invasive bacterial pathogens in resource-poor settings, and cause significant mortality, particularly in children. But neither of the two available vaccines can be used in infants. Interestingly, Vi conjugate vaccines should offer children excellent protection from typhoid [24].

There is evidence that Ty21a vaccine may have cross-protective efficacy against numerous non typhoidal *Salmonella enteritidis and typhimurium* [9]. Greater effort must be placed on the development of next generation vaccines to address the disease burden resulting from typhi, paratyphi A and non-typhoidal Salmonella NTS infections [25]. Policy-makers therefore should introduce preventive interventions against typhoid, including vaccination [26].

5. CONCLUSION

Typhoid carrier rate in Enugu is taken as 2.1% while that of paratyphoid salmonella carrier rate is 1.0 percent. From the study females with a raised Widal titre160 and above to 'H' antigen are more likely to be typhoid carriers in Enugu. Provision of potable water, proper treatment and vaccination should lower the typhoid carrier rate in the community.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- 1. Parry CM, Wijedoru I, Arjyal A, Bakers. The utility of diagnostic tests for enteric fever in endemic locations. Expert Rev Anti Infect Ther. 2011;9:711-725.
- 2. Hook E. W Salmonella species (including typhoid fever). In Mandell G. L, Douglas R.G, Jnr Bennett J. E eds. Principles and Practice of infectious diseases. Churchill Livingstone. 1990;1700-1716.
- 3. Olubuyide IO. Typhoid fever in the tropics post grad doctor Africa. 1992;14:37-41.

- Giannella RA. In Medical Microbiology. Baron S. edit 3rd ed. Churchill Livingstone New York. 1991;317–325.
- 5. Mandel GL, Douglas RG, Benneth JE. Principles and practice of infectious diseases. 3rd ed. Churchill Livingstone New York. 1990;1700–1716.
- Ferreccio C, Levine M, Astroza L, Berrios G, Solari V, Misraji A, Petaur C. The detection of chronic *Salmonella typhi* carriers: A practical method applied to food handlers. Rev Med Chil. 1990;118:33–37.
- Onuigbo MAC. Diagnosis of typhoid fever in Nigeria: Misuse of the Widal test. Trans Roy. Soc. Trop Med Hyg. 1990;84(1):129-131.
- Vanishnavi C, Kochhar R, Singh G, Kumar S, Singh S, Singh K. Epidemiology of typhoid carriers among blood donors and patients with biliary, gastrointestinal and other related diseases. Microbiology and Immunology. 2005;49(2):107-112.
- Kantele A, Pakkanem SH, Siitonen A, Karttunen R, Kantele JM. Live oral typhoid vaccine Salmonella typhi Ty21a – a surrogate vaccine against nontyphoid Salmonella? Vaccine. 2012;30(50):7238– 45.
- Kantele A, Parkkanen SH, Karttunem R, Kantele JM. Head –to-head comparison of humoral immune responses to vacapsular polysaccharide and *Salmonella typhi* Ty21a typhoid vaccines – a randomized trial. Plos One. 2013;8(4):e60583.
- 11. Pakkannen SH, Kantele JM, Herzog C, Kantele A. Cross-reactive immune response elicited by parenteral Vi polysaccharide typhoid vaccine against non-typhoid Salmonellae. Vaccine. 2014;32(5):5444-51.
- 12. Hawkey PM, Lewis DA. Medical bacteriology, a practical approach 2nd ed. Oxford University Press United Kingdom. 1994;21–42.
- Ismail A. New advances in the diagnosis of typhoid and defection of typhoid carriers. Malays J. Med. Sci. 2000;7(2):3-8.
- 14. Dutta P, Mitrall, Dtta S, Mitra U, Dutta S, De A, Chatterjee MK, Bhattacharya SK. Ceftriaxone therapy in ciprofloxacin treatment failure typhoid fever in children. Indian J. Med. Res. 2001;113:210-213.
- 15. Effa EE, Bukirwa H. Arithromycin for treating uncomplicated typhoid and paratyphoid fever (enteric fever). Cochrane Data base of systematic Reviews. 2008;8(4):CD006083

- Finkbeiner AK. Quite contrary: Was "Typhoid Mary" Mallon a symbol of the threats to individual liberty or a necessary sacrifice to public health. The sciences. 1996;36(5):38-43.
- 17. Ukwenya AY, Ahmed A, Garba ES. Progress in management of typhoid perforation. Ann Afr Med. 2011;10(4):259-65.
- Dutta U, Gorg PK, Kumar R, Tandon RK. Typhoid carriers among patients with gallstones are at increased risk for carcinoma of the gallbladder. Am J gastroenterol. 2000;95:784-787.
- Khatri NS, Maskey P, Pondel S, Jaiswal VK, Karkey A, Koirala S, Shakya N, Agrawal K, Arjyal A, Basnyat B. Gallbladder carriage of Salmonella paratyphi A may be an important factor in the increasing incidence of this infection in South Asia. Ann Intern Med. 2009;150:567–569.
- 20. Karkey A, Aryjal A, Basynyat B, Baker S. Kathmandu, Nepal; Still an enteric fever capital of the world. JIDC. 2008;2:461– 465.
- 21. Crump J A, Luby S P, Mintz E D. The global burden of typhoid fever. Bull World Health Organ. 2004;82(5):346–353.

- 22. Typhoid vaccines: WHO position paper. Wkly Epidemiol. Rec. 2008;83(6):49–59.
- 23. Parkkanen SH, Kantele JM, Kantele A. Cross-reactive gut-directed immune response against *Salmonella enterica serovar paratyphi* A and B in typhoid fever and after oral Ty21a typhoid vaccination. Vaccine. 2012;30(42):6047–53.
- 24. MacGregor AC, Waddinton CS, Polland AJ. Prospects for prevention of Salmonella infection in children through vaccination. Curr Opin Infect Dis. 2013;26(3):254–62.
- 25. Martin LB. Vaccines for typhoid fever and other Salmonellases. Curr Opin Infect Dis. 2012;25(5):489–99.
- 26. R Leon Ochiai, Camilo J Acosta, M Carolina Danovaro-Holliday, Dong Baiqing, Sujit K Bhattacharya, Magdarina D Agtini, Zulfiqar A Bhutta, Do Gia Canh, Mohammad Ali, Seonghye Shin, John Wain, Anne-Laure Page, M John Albert, Jeremy Farrar, Remon Abu-Elyazeed, Tikki Pang, Claudia M Galindo, Lorenz von Seidlein, John D Clemens, A study of typhoid fever in five Asian countries: Disease burden and implications for control. Bull World Health Organization. 2008;86(4):260–268.

© 2014 Ohanu and Ogeneh; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

> Peer-review history: The peer review history for this paper can be accessed here: http://www.sciencedomain.org/review-history.php?iid=657&id=8&aid=6039