

Journal of Pharmaceutical Research International

33(60B): 3480-3486, 2021; Article no.JPRI.80385 ISSN: 2456-9119

(Past name: British Journal of Pharmaceutical Research, Past ISSN: 2231-2919, NLM ID: 101631759)

Scaffolds in Periodontal Regeneration

Rohan Khetan a^{*≡}, Khushbhoo Durge bo, Pavan Bajaj b#, Bhairavi Kale bo and Nutan Dhamdhere at

^a Sharad Pawar Dental College and Hospital, Datta Meghe Institute of Medical Sciences (Deemed to be University), Sawangi (Meghe) Wardha.442001, Maharashtra, India. ^b Department of Periodontics, Sharad Pawar Dental College, Datta Meghe Institute of Medical Sciences (Deemed to be University), Sawangi (Meghe) Wardha.442001, Maharashtra, India.

Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/JPRI/2021/v33i60B35036

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: https://www.sdiarticle5.com/review-history/80385

> Received 22 October 2021 Accepted 26 December 2021 Published 28 December 2021

Review Article

ABSTRACT

The periodontium is a well-designed central core made up of numerous tissues that surround and support the tooth. Sustained periodontitis has the ability to create havoc on periodontal tissues. leading to tooth loss. Diverse biomaterials are used as obstacle layers in the coordinated-tissuerecuperation (GTR), which at present is the highest quality level under the dentistry community, to treat periodontally weak teeth. Unique biomaterials have recently been structured in a tissue planning stage to aid in the recovery of injured periodontal tissues. In recent decades, innovations in stage manufacture have ranged from a genuine substrate to aid in the recovery of a specific kind of periodontal tissue to a multiphase/bioactive stage structure to coordinate a coordinated periodontal tissue recovery. This article discusses the new sorts of phrases that are being developed for periodontal tissue recovery.

Keywords: Cementum; gingival; periodontal ligament; periodontal; periodontium; periodontitis; regeneration; scaffolds, syndrome torrent.

Third BDS Student:

^ω Assistant lecturer;

[#] Associate Professor;

[†] Final BDS Student;

^{*}Corresponding author: E-mail: rohankhetan7415@gmail.com;

1. INTRODUCTION

Oral diseases are the most common cause of periodontal disease. lf the periodontal ligament, cementum, bone, alveolar gingiva are not treated, microscopic organisms that cause aggravation could spread and ruin them. Periodontitis is diagnosed when x-rays reveal Alveolar bone destruction is a condition under which the alveolar bone is eliminated, and it is usually believed to be an irreversible condition. Gingivitis is non-medication-required chronic gum disease. As a result, gingival recession is frequently connected with functional and cosmetic issues such as root caries and black triangles. In addition to cosmetic and functional concerns, periodontitis is associated with systemic illnesses including congestive heart failure, diabetes, respiratory disease, etc [1]. As a result, gingivitis is becoming a serious general public health problem in which finding effective periodontal therapies should be a key focus for health researchers. The frequency of mild to moderate periodontitis was highest in the urban population (21.7%). (CI: 16.3-27.5). Females exhibited a lower frequency of periodontitis (33.4%) than males (43.2%).

As per by the National Health & Nutrition Examination Survey (NHANES), nearby 7.9% of grown people (ages 18 to 62) and 18.6% of seniors (ages 60-70) in the United States have Periodontitis, while tooth caries affects 39% of children (ages 3 to 9) in their baby tooth and 60% of growing people (ages 11 to 21) in there fix the tooth. Two disorders continue to be a major public health issue that requires more effective preventive and management techniques [2].

2. OBJECTIVE OF PERIODONTAL REGENERATION VIA BIOMATERIAL SCAFFOLDS

2.1 Ancient Legacy of Periodontal Regeneration

The focusing aim for therapy in periodontitis is to recreate periodontal tissues that work together, in which cementum, PDL, and AB fibers aligned longitudinally between CM and AB and CM, PDL, and AB are created synchronously in their proper places. The bacterial infection causes periodontitis, which seems characterized by increased neutrophil and macrophage infiltration, osteoclast activation via RANKL signaling, and bone resorption [3]. Plaque and calculus on the

teeth increase the risk of bacterial infection and periodontitis by creating a space between the teeth and the periodontium [4]. Non-surgical, conservative therapy that addresses the causes of gingivitis (e.g, calculus and dental plaque).

Tissue engineering technologies for periodontal regeneration have lately been studied as a novel way to overcome such limitations [5]. GTR can be used with a variety of biomaterial scaffolds that include cells and/or bioactive. More improved scaffold technologies for guiding integrated periodontal regeneration have recently been developed [6]. Until they degrade and are replaced by new tissues, these scaffolds are meant to offer bioactive indicators for periodontal regeneration.

2.2 Scaffolds to Aid in Periodontal Wound Repair

In tissue engineering, scaffolds are utilised to create a platform for cell adhesion, tissue ingrowth, and early structural support [7]. Nondegradable or degradable membranes impede formation GTR. epithelium in periodontal connective tissue and PDL to heal slowly (4-6 weeks) [8]. A long period of periodontitis, on the other hand, may aggravate the result of GTR by decreasing PDL cell repairability, hindering the host immunity, or significantly denaturalizing the cememtum [9]. Periodontal tissue regeneration scaffolds may provide contact guidance, allowing cells to move more quickly into periodontal abnormalities and speed up periodontal tissue regeneration scaffolds [10].

Scaffold systems have proved to offer a lot of promise for periodontal regeneration. The current technological advancement in micro precise regional under scaffold designing is marked as an essential step toward integrated multi tissue periodontium regeneration. There are numerous scaffolds and delivery systems used to regenerate integrated periodontium.

2.3 Scaffolds for Periodontal Regeneration have an Antimicrobial Impact

Bacteria are the significant cause of periodontitis, and the ailment is triggered by the host's defensive resistance to them [11]. Despite the fact that modern periodontal therapy involves eliminating dental biofilm and calculation, which are the main sources as well as niches of microbes that produce clinical indicators. periodontitis reoccurrence is tightly tied to maintenance to avoid secondary infection [12]. Current periodontal treatments provide a risk of bacterial re-infection. As a result, antimicrobial activity has been included in various scaffolds meant to improve periodontal regeneration while also lowering the likelihood of re-infection following periodontal treatment. Antibacterial material components have been incorporated scaffold materials in a simple straightforward manner. Chitosan is a bioadhesive, antifungal, antibacterial, and hemostatic polymer generated from natural seashells [13].

2.4 Scaffolds for Periodontal Regeneration with Anti-inflammatory Characteristics

Periodontitis is a multifaceted inflammatory condition, an illness that causes the periodontium to deteriorate over time [14]. Despite the fact that bacterial infection appears to be the primary cause of periodontitis, Periodontitis is triggered by an overactive immunological response from the host and/or an absence of inflammation pharmacological resolution. As а result. medications which include NSAIDS medicines which are used to try to reduce inflammation [15]. Treatments that target just proinflammatory pathways and /or signaling without addressing causative factors, on the other hand, have minimal benefits [16]. Only as a supplement are they used. The removal of the causative causes (such as dental biofilm as well as calculus), the initial stage in periodontal treatment is to remove inflammatory granulation tissue.

3. BIOMATERIALS FOR PERIODONTAL REGENERATION SCAFFOLD (TOTAL A1)

3.1 Natural Materials

Cell affinity and biocompatibility are often high in natural biomaterials. They are less toxic and elicit inflammatory and immunological reactions less frequently. As a result, natural biomaterials have become popular periodontal tissue as regeneration scaffolds [17]. Βy its immunogenicity, biodegradability, biocompatibility, biodegradability, and antibacterial capability as opposed to fungus and bacteria. chitosan has been frequently employed

for periodontal regeneration [18]. Chitosan is typically combined with other scaffold materials which can provide antimicrobial action due to its drawbacks as standalone scaffold material, it includes particle aggregation as well as limited solubility [19].

3.2 Bioceramics

Bioceramic scaffolds are ideal for the regeneration of the periodontium because of their mechanical durability and biodegradability [20]. The key benefits of bio-ceramic-based scaffolds over all the other natural and synthetic materials excellent osteoconductive their osteoinductive properties [21]. Bioceramics can be used to treat periodontal disorders and come in a variety of forms, including granules, pastes, and injectables [22]. Bio-active glass, a popular bioceramic having high osteogenesis qualities that are often used for periodontal tissue regeneration, is another well-known bioceramic with high osteogenic capabilities [23].

3.3 Synthetic Polymers

To replace the non-resorbable PTFE membrane. synthetic polymers have been predominantly 2nd employed as Generation degradable membrane materials. Many polyester-based polymers have been employed as periodontal scaffold materials [24]. Synthetic polymers offer some advantages, including highly variable physio-chemical capabilities. variable biodegradation rates. and low-cost. conventional manufacturing technology that enables mass production [25].

4. TISSUE ENGINEERING FOR PERIODONTAL REGENERATION RECENT PROGRESS

4.1 Scaffolds having Intelligence

Thanks to recent breakthroughs in polymer engineering, functional scaffolds have been constructed. Cell culture surface modification or polymer design can alter cell orientation and kind rhPDGF- BB [26]. was used to construct a 3D printed bioresorbable scaffold for periodontal regeneration. The treated area remained intact for 11 to 12 months after the therapy, according to the authors [27].

4.2 Scaffold-free Culture Technique

A spheroid culture strategy was devised for PDL-MSCs with improved osteogenic capacity, and

this technique may be effective for hard tissue regeneration [28]. Another study discovered that spheroid culture altered the expression of anti-inflammation and angiogenesis genes in PDLSCs. These alterations are thought to be linked to apoptosis signaling [29]. These data suggest that culture conditions affect the characteristics of PDL-MSCs.

4.3 Tissue-engineered Constructs that have been Decellularized

Decellularized materials have as of late been made for periodontal recovery. Chemical decellularization of HPDL cell sheet with DNase and NH4OH/Triton X-100 resulted in positive extracellular matrices are intact, and growth factors are accessible, which can lead to promoting allogeneic cell proliferate [30]. Before being implanted into rat periodontal insufficiency models [31]. The absence of essential cells in the decellularized construct aided attachment formation, implying that the construct itself aided periodontal regeneration. As a result, this product may be authorized to be acquired off the shelf [32].

5. FUTURE CYTOTHERAPY FOR PERIODONTAL REGENERATION:- A PERSPECTIVE

5.1 Cell Bank Establishment and Allogeneic Cell Sources

PDL-MSC sheets were found to be safe and feasible for therapeutic application in autologous transplantation research [33]. However, some of the strategy's flaws have become apparent. For starters, individuals without redundant teeth are unable to get this treatment due to a lack of a cell supply almost all older people seem to be missing wisdom teeth, that are most likely extracted when they were younger Cell growth and phenotypic differences between individuals are seen in certain patients with duplicated teeth. As a result, it's challenging to make things that are both reliable and stable [34]. This cellular design is produced from the tooth and takes four weeks to generate, making it an expensive technique. As a result, we made a decision to modify our opinions.

5.2 ES/ IPS Cells (Embryonic Stem/ Induced Pluripotent Stem)

Furthermore method to establish a cell bank is to IPS /ES cells, which are projected to reproduce

indefinitely. Overexpression of 4 key factors of transcription is c-Myc, Sox2, Klf4, Oct4, and can be used to give rise to iPS cells from somatic cells that resemble ES cells. Despite the fact that the approach and methodology for producing PDL cells are unclear, a strategy for generating MSCs from ES/iPS cells utilizing neural crest cells has been created [35]. PDL cells are derived from dental follicle cells development [36]. As a result, recognizing the economic growth from dental follicles to PDL tissues and cells is critical. MSC growth can be modified by the stiffness of cultureware. according to recent studies [37].

5.3 MSCs' Condition Medium

According to a recent study, MSCs generate and release proteins and extracellular vesicles into the culture medium, and conditioned media possesses regeneration potential. Nagataetal collected and condensed PDL-MSC conditioned media to mimic periodontal deficits in rats [38]. TNF mRNA levels were also lower in periodontal tissues treated with PDL-MSC conditioned medium. According to these findings, PDL-MSC's conditioned medium improved regeneration of the periodontitis by reducing inflammatory responses via TNF production. Although the specific mechanisms are unknown, current research suggests that in addition to conditioned media proteins. extracellular vesicles (EVs) that carry proteins, mRNAs, mi RNAs, and DNA. The EV fractions from the conditioned medium were the focus of our research [39].

6. CONCLUSION

The cell—scaffold interaction that might also occur when tissue-engineering technologies are adopted for regeneration of periodontitis was the center of this review. Biomaterial treatments can still be used in dental clinics thanks to recent innovations periodontal regeneration in technology. Whether stem cells are being used not, the biocompatible scaffold is another key technique in tissue engineering technology that has been embraced for the construction of new tissue [40]. Recent developments in the design and fabrication of excellent biomaterial scaffolds are a massive breakthrough in periodontal regeneration utilizing tissue engineering technologies. The periodontium is a complex multi-tissue structure, and its renewal is inextricably related to periodontitis' severe / chronic inflammation.

Several newer hvdrogel-based delivery technologies also allowed for said timely and/or stimulation-sensitive release of numerous components that could be included under different scaffold architectures [41]. We spoke about the prospective periodontal cell- scaffold interactions, which might be a helpful technique for searching for the perfect scaffold. Many cellular and molecular processes involved in periodontal tissue healing systems have been discovered to date. These breakthroughs in knowledge might be the catalyst for a breakthrough in cell-based regeneration techniques in our profession. Rapid advances in material sciences and micro/nanofabrication technologies are also paving the door for novel tissue engineering approaches for "perfect" periodontal regeneration.

CONSENT

It is not applicable.

ETHICAL APPROVAL

It is not applicable.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- 1. Pihlstrom BL, Michalowicz BS, Johnson NW. Periodontal diseases. Lancet. 2005;366:1809–1820. [CrossRef]
- Bottino MC, Thomas V, Schmidt G, et al. Recent advances in the development of GTR/GBR membranes for periodontal regeneration--a materials perspective. Dent Mater. 2012; 28(7):703–721. [PubMed: 22592164]
- 3. Z. Feng, A. Weinberg, Role of bacteria in health and disease of periodontal tissues, Periodontology. 2006;40(1):50–76, 2000.
- 4. Lindhe J, Nyman S. The effect of plaque control and surgical pocket elimination on the establishment and maintenance of periodontal health. A longitudinal study of periodontal therapy in cases of advanced disease, J. Clin. Periodontol. 1975;2(2):67–79.
- 5. Mishra M, et al. Scaffolds in periodontal regeneration, J. Pharmaceut. Biomed. Sci. 2016;6:10–16.

- 6. Pilipchuk SP, et al. Micropatterned scaffolds with immobilized growth factor genes regenerate bone and periodontal ligament-like tissues, Adv. Healthcare Mater. 2018;7(22):180-0750.
- 7. Sari DS, et al. Osteogenic differentiation and biocompatibility of bovine teeth scaffold with rat adipose-derived mesenchymal stem cells, Eur. J. Dermatol. 2019;13(2):206–212.
- Cortellini P, et al., Periodontal regeneration of human infrabony defects. I. Clinical measures, J. Periodontol. 1993;64(4):254– 260.
- 9. Zheng W, et al. Periodontitis promotes the proliferation and suppresses the differentiation potential of human periodontal ligament stem cells, Int. J. Mol. Med. 2015;36(4):915–922.
- D. Carmagnola, et al., Engineered scaffolds and cell-based therapy for periodontal regeneration, J. Appl. Biomater. Funct. Mater. 2017;15(4):0-0.
- 11. Lee JH, et al. Adjunctive use of enamel matrix derivatives to porcine-derived xenograft for the treatment of one-wall intrabony defects: two-year longitudinal results of a randomized controlled clinical trial, J. Periodontol. 2020;91(7):880–889.
- Darveau RP, Periodontitis: A polymicrobial disruption of host homeostasis, Nat. Rev. Microbiol. 2010;8(7):481–490.
- 13. Rana D, Sande S. Study of Prevalence and Antimicrobial Susceptibility Pattern of Enterococci Isolated from Clinically Relevant Samples with Special Reference to High Level Aminoglycoside Resistance (HLAR) in a Rural Tertiary Care Hospital. Journal of Evolution of Medical and Dental Sciences. 2020;9(34):2472-9.
- Ramfjord SP. Maintenance care for treated periodontitis patients, J. Clin. Periodontol. 1987;14(8):433–437.
- 15. Abdal-Wahab M, et al. Regenerative potential of cultured gingival fibroblasts in treatment of periodontal intrabony defects (randomized clinical and biochemical trial), J. Periodontal. Res. 2020;55(3):441–452.
- Papapanou PN, et al. Periodontitis: consensus report of workgroup 2 of the 2017 world workshop on the classification of periodontal and peri-implant diseases and conditions, J. Periodontol. 2018;89(Suppl 1):S173–s182.
- 17. Van Dyke TE. The management of inflammation in periodontal disease, J. Periodontol. 2008;79:1601–1608.

- Ashworth JC, et al. Optimising collagen scaffold architecture for enhanced periodontal ligament fibroblast migration, J. Mater. Sci. Mater. Med. 2018; 29(11):166.
- Zang S, et al. Injectable chitosan/βglycerophosphate hydrogels with sustained release of BMP-7 and ornidazole in periodontal wound healing of class III furcation defects, Mater. Sci. Eng. C 2019;99:919–928.
- 20. Liao Y, et al., Mesoporous hydroxyapatite/chitosan loaded with recombinanthuman amelogenin could enhance antibacterial effect and promote periodontal regeneration, Front. Cell. Infect. Microbiol. 2020;10.
- 21. Chen Q, et al. Progress and challenges in biomaterials used for bone tissue engineering: bioactive glasses and elastomeric composites, Prog. Biomater. 2012;1(1):2.
- 22. Ghassemi T, et al., Current concepts in scaffolding for bone tissue e ngineering, Arch. Bone Joint Surg. 2018;6(2):90.
- 23. Laschke MW, et al. Injectable nanocrystalline hydroxyapatite paste for bone substitution: *In vivo* Analysis of Biocompatibility and Vascularization. 2007;82B(2):494–505.
- 24. Amin AMM, Ewais EMM, Bioceramic Scaffolds, In Tech; 2017.
- 25. Nagy K, et al. A novel hydrogel scaffold for periodontal ligament stem cells, Interv Med Appl Sci 2018;10(3):162–170.
- 26. Carter SSD, et al. Additive biomanufacturing: an advanced approach for periodontal tissue regeneration, Ann. Biomed. Eng. 2017;45(1):12–22
- 27. Zhang L, et al. Three-dimensional (3D) printed scaffold and material selection for bone repair, Acta Biomater. 2019;84: 16–33.
- 28. Sethiya KR, Dhadse PV. Healing after Periodontal Surgery--A Review. Journal of Evolution of Medical and Dental Sciences. 2020;9(49):3753-60
- A.P. 29. Nugraha, et al., Gingival mesenchymal stem cells and chitosan scaffold to accelerate alveolar bone remodelling in periodontitis: a narrative review, Res. J. Pharm. Technol. 2020;13(5):2502-2506.
- Ammar MM, et al. Growth factor release and enhanced encapsulated periodontal stem cells viability by freeze-dried platelet

- concentrate loaded thermo-sensitive hydrogel for periodontal regeneration, Saudi Dental J. 2018;30(4): 355–364.
- Farag A, Vaquette C, Theodoropoulos C, 31. Hamlet SM, Hutmacher DW, Ivanovski S. periodontal Decellularized ligament cell sheets with recellularization potential. Dent. Res. 2014;93: J. 1313-1319.
- 32. Panchbhai A. Nanocomposites: Past, present, and future of dentistry. InApplications of Nanocomposite Materials in Dentistry. Woodhead Publishing. 2019:181-190.
- 33. Wankhede AN, Dhadse PV. Role of Interleukin-17 in immunopathology of chronic and aggressive periodontitis. Journal of the International Clinical Dental Research Organization. 2019;11(1):3.
- 34. Farag A, Hashimi SM, Vaquette C, Bartold PM, Hutmacher DW, Ivanovski S. The effect of decellularized tissue engineered constructs on periodontal regeneration. J. Clin. Periodontol. 2018;45: 586–596.
- 35. Iwata T, Yamato M, Washio K, Yoshida T, Tsumanuma Y, Yamada A, Onizuka S, Izumi Y, Ando T, Okano T et al. Periodontal regeneration with autologous periodontal ligament-derived cell sheets— A safety and efficacy study in ten patients. Regen. Ther. 2018;9:38–44.
- 36. Fleischmannova J, Matalova E, Sharpe PT, Misek I, Radlanski RJ. Formation of the tooth-bone interface. J. Dent. Res. 2010;89:108–115.
- 37. Lv H, Li L, Sun M, Zhang Y, Chen L, Rong Y, Li Y. Mechanism of regulation of stem cell differentiation by matrix stiffness. Stem Cell Res. Ther. 2015;6: 103.
- 38. Nagata M. Iwasaki K, Akazawa K, Yokoyama N, Izumi Y, Komak M. Morita I. Conditioned Medium from Periodontal Stem Ligament Cells Enhances Periodontal Regeneration. Tissue Eng. Part A. 2017;23: 367-377.
- 39. Sjoqvist S, Ishikawa T, Shimura D, Kasai Y, Imafuku A, Bou-Ghannam S, Iwata T, Kanai N. Exosomes derived from clinical-grade oral mucosal epithelial cell sheets promote wound healing. J. Extracell. Vesicles 2019;8: 1565264.

- 40. Garg T, Singh O, Arora S, Murthy R. Scaffold: a novel carrier forcell and drug delivery. Crit Rev Ther Drug Carrier Syst H. Shimauchi et al. 2012;29:1 63.130
- 41. L.L. Wang, et al., Injectable and protease-degradable hydrogel for siRNA sequestration and triggered delivery to the heart, J. Contr. Release. 2018;285:152–161.

© 2021 Khetan et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:
The peer review history for this paper can be accessed here:
https://www.sdiarticle5.com/review-history/80385