



Clinical Significance of Mir-221 in Colorectal Cancer

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Authors' contributions

This work was carried out in collaboration among all authors. The Initial manuscript was prepared by author WHE, the study was conceptualized and designed by authors AFG, ARNA and AEA. The final data preparation was done by authors AFG, WHE and MNA and the statistical analysis completed by author WHE and final data analysis and interpretation finally assembled by authors WHE, AFG, AEA. All the contributing authors have taken part in manuscript editing and review.

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ABSTRACT

Aim: MicroRNA-221 plays an important role in a number of human malignancies. The present study was conducted to shed light on the role of mir-221 in colorectal cancer as regards the initiation of malignant process and progression.

Methods: Real-time reverse transcription–polymerase chain reaction was used to determine the levels of mir-221 in 92 patients with colorectal cancer and in their adjacent non-cancerous tissues and to explore the relation between mir-221 level and clinical and pathological features of the disease.

Results: Mir-221 was up-regulated in 90.2% (83/92) of colorectal cancer tissue samples compared with their adjacent non-cancerous tissue samples. The high expression of mir-221 was significantly correlated to tumor size and infiltration, clinical stage and lymph node metastasis. Univariate and multivariate analyses showed that over expression of mir-221 was an unfavorable prognostic indicator for overall survival in colorectal cancer patients.

Conclusion: Our findings suggest mir-221 may be used as a prognostic marker in CRC and it needs more studies to be used as a potential diagnostic marker and as a molecular target for targeted cancer therapy.

Keywords: Colorectal cancer; mir-221; RT-PCR.

1. INTRODUCTION

Colorectal cancer (CRC) is the third prevalent cancer type worldwide and the main cause of mortality from malignant tumors in both sexes [1,2]. In spite of the fact that CRC incidence has been decreasing due to decreased exposure to social risk factors, the five years survival rate is unsatisfactory 64.9% necessitating the need for early detection [3,4].

MiRNA is a kind of endogenous small RNA with a length of around 25 nucleotides, which plays various important roles in early development, fat metabolism, cell differentiation and other biological activities [5]. The expression of miRNAs is closely related to the occurrence and development of human malignancies. miRNAs participate in the proliferation, apoptosis, and invasion of tumor cells either as anti-oncogenes or oncogenes [6].

MicroRNA-221 (miR-221), encoded on human chromosome X, is overexpressed in many aggressive carcinomas [7-9]. Abnormal overexpression of miR-221 strongly facilitates tumor cell growth by inducing cell lines in vitro to progress into the S phase of the cell cycle [10].

Several studies have discovered that miR-221 is significantly up-regulated in cell lines [11], plasma or serum [12-15], and tissues of many human malignancies [16-20].

Elevated expression of miR-221 in many types of carcinomas is obviously related to an easier tumor invasion [10,17,21-23], a larger size of tumor [13,17,23] earlier distant metastasis [17,22,24], and shorter time to recurrence and shorter survival [24-26].

Studies have revealed a close association between the level of miR-221 expression and clinic pathological tumor features including tumor-node-metastasis (TNM) stage, local

invasion, metastasis, prognosis, radiosensitivity and anticancer drug resistance [22,27] but the role of miR-221 in colorectal progression and metastasis is still unclear.

The current study was designed to compare miR-221 expression in colorectal cancer tissues and adjacent normal tissues, to study the prognostic significance of miR-221 expression and its relation to the clinic pathological characteristics of the tumors.

2. PATIENTS AND METHODS

Tissue samples from primary colorectal cancer and adjacent non-cancerous tissue at least 3 cm from tumorous tissue were obtained surgically from 92 patients with histo-pathologically proven colorectal cancer, who underwent curative surgical resections at colorectal cancer unit, Faculty of Medicine, Zagazig University between June 2012 and September 2017. All patients with previous malignancy or with previous chemotherapy, immunotherapy or radiation therapy were excluded from the study.

After surgery tissues were flash-frozen and stored at -80°C in liquid nitrogen. Patients were staged according to the American Joint Committee on Cancer (8th edition) [28]. The study was approved with the medical ethics committee of our institution. Informed consent was obtained from all individual participants included in the study.

2.1 RNA Extraction and Quantitative Real-Time RT-PCR

TRIzol® reagent (Invitrogen, Carlsbad, CA, USA) was used according to the manufacturer's instructions, to isolate RNA from 20-30 mg of colorectal samples and their adjacent non-cancerous normal tissues. The RNA quantity and quality was assessed by Bioanalyzer 2100 system (Agilent Technologies, Santa Clara, CA, USA).

TaqMan® miRNA (Applied Biosystems, Foster City, CA, USA) and specific gene primers were used to synthesize the complementary DNA of MiR-221 and RNA U6 (internal control) from total RNA.

Reverse transcription (RT) primer sequences (Invitrogen) were 5'-GTCGTATCCAGTGCAGGGTCCGAGGTATTCGACTGGATACGACGAAACCCA-3' (for miR-221) and 5'-CGTTCACGAATTTGCGTGTCAT-3' (for RNA U6). The reaction mixture of RT consisted of 50 nmol / l RT primer, 1 × RT buffer, 3.33 U/μl MultiScribe® reverse transcriptase, 10 ng total RNA, and 0.25 mmol / l of deoxynucleotide triphosphate and 0.25 U/μl RNase inhibitor in a total volume of 7.5 μl (Purchased from Applied Biosystems). Reactions were incubated in a 96-well plate for 30 min at 16°C, followed by 30 min at 42°C and 5 min at 85°C, followed by 4°C.

Reactions were carried out in 96-well plates in a total volume of 10 μl (0.67 μl RT products, 1 × TaqMan® Universal PCR master mix, and 1 μl TaqMan® miRNA assay primer and probe mix). Real-time polymerase chain reaction (PCR) was performed using the 7500 real-time PCR system (Applied Biosystems).

The program of cycling consists of preliminary denaturation at 95°C for 10 min, followed by 45 cycles of denaturation at 95°C for 15 s, annealing at 55°C for 30 s and elongation at 70°C for 30 s, followed by a final elongation step at 60°C for 10 min.

All samples were processed in triplicate. When the fluorescence passed a fixed threshold, this cycle number is defined as the threshold cycle (CT). The relative amount of miR-221 to U6 was calculated using equation $2^{-\Delta CT}$,

$$\text{where, } \Delta CT = (C_{T \text{ miR-221}} - C_{T \text{ U6}}). [29]$$

2.2 Statistical Analysis

Statistical analysis was performed with SPSS® version 23.0 (SPSS Inc., Chicago, IL, USA) for Windows®. Data were expressed as mean ± SD. Differences in expression levels of miR-221 between groups were compared using Student's *t*-test. Kaplan–Meier curves were used to calculate the survival of the patients and the curves were compared by the log-rank test. The Cox proportional hazards regression model was used to examine the joint effect of co-variables.

3. RESULTS

Tissues samples obtained by surgery from 92 patients with colorectal cancers and another sample obtained from normal tissues adjacent to the tumor and located at least 3-5 cm from the tumor were obtained for each patient. Our study consisted of 49 Males and 43 females with a mean age ± SD (52.5 ± 6.4). MiR-221 was up-regulated in 90.2% (83/92) of colorectal cancer tissue samples compared with their adjacent non-cancerous tissue samples. The mean ΔCT value of mir-221 ± SD in colorectal cancer samples was 2.46 ± 0.32, compared with 0.86 ± 0.14 in their adjacent non-cancerous tissue samples ($P < .001$). The correlations between the clinical and pathological data of patients and miR-221 expression levels (Table 1).

Advanced Clinical stage III ($P = .018$), invasion of the tumor (T) ($P = .006$) and lymph nodes metastasis ($P = .032$), all showed significant correlation with high mir-221 expression. Meanwhile, no significant correlation was noticed between age, gender, and degree of tumor differentiation and level of mir-221 expression.

At the time of survival analysis (March 2019), fifteen patients out of ninety-two (16.3%) are still alive. The range of follow-up period was (11- 65 months) and the median duration of follow-up was 42 months. During follow-up distant metastasis was observed 52 patients as the first recurrence, 16 patients had a local recurrence, while both distant metastasis and local recurrence was noted in 4 patients as the first relapse.

According to the mean level of mir-221 (2.46 ΔCT), patients were divided into patients with low mir-221 expression (n=44) and those with high expression (n = 48). The survival between low and high expression groups is shown in Fig. 1. The 4-year survival rates were 44% in patients with low expression and 35% in those with high expression with corresponding median survival times of 41 and 35 months respectively ($P = 0.016$).

The univariate analysis revealed that patients the female gender having worse prognosis; (hazard ratio [HR] 0.435; $P = 0.23$), high-level miR-221 expression (HR 2.422; $P = 0.016$), tumor size and infiltration (T) (HR 5.652; $P < 0.001$), tumor stage (TNM) (HR = 3.462; $P = 0.008$), lymph node metastasis (HR 2.956; $P = 0.006$) and

degree of tumor differentiation (HR 5.165; $P = 0.011$) all were predictive factors for prognosis in patients with colorectal cancer (Table 2).

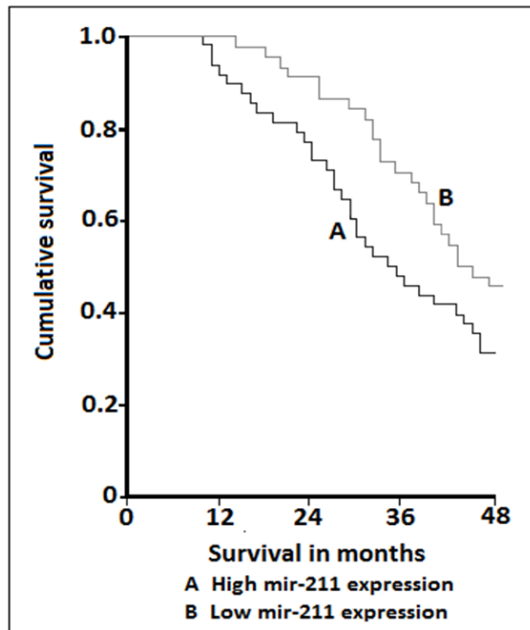


Fig. 1. Kaplan–Meier survival curve for patients with colorectal cancer; the 4-year survival rate and the median survival time were significantly lower in patients with high miR-221 expression compared with patients with low miR-221 expression ($P = 0.016$, both comparisons; log-rank test)

Multivariate Cox regression analyses revealed that the high level of mir-221 expression was the only unfavorable factor, independent of other factors as tumor size and infiltration (T) (HR 4.268; $P < 0.001$), tumor stage (TNM) (HR 3.225; $P = 0.016$), lymph node metastasis (HR 2.685; $P = 0.026$) and degree of tumor differentiation (HR 3.678; $P = 0.032$) (Table 3).

4. DISCUSSION

Colorectal cancer (CRC) is one of the most common cancers worldwide, accounting for 10% of all new cancer cases and over 1.23 million deaths per year [1].

Early-stage CRC usually has a favorable prognosis but unfortunately about two-thirds of patients present with locally advanced or metastatic disease [30].

Significant progress in molecular pathogenesis and treatment of CRC has been made in the last decade, but advanced stages CRC is still difficult

to treat because CRC is a group of diverse heterogeneous diseases arising through various molecular pathways. The prognosis and response to treatment are determined by this heterogeneity that made a big challenge to study the molecular basis of the disease. To date, there are a few molecular markers as KRAS, microsatellite instability, the mutation status of BRAF and PICK3CA that are used in treatment decisions and in clinical patient management. To better define CRC patients with aggressive and lethal disease, we need to identify new molecular biomarkers.

The understanding of gene regulation has been dramatically changed since the discovery of miRNAs; research findings have emphasized the important role of miRNAs in the molecular biology of cancer.

miRNAs are a group of small non-coding RNA molecules that regulate gene expression at the post-transcriptional level. There are more than 2500 miRNAs have been identified in humans (miRBase database 20.0) [31].

In particular, miR-221 has been demonstrated to be an important oncogenic miR in many cancers [12–21], but the associations between miR-221 expression levels, clinicopathological characteristics, and survival in patients with gastric cancer remain unclear, particularly has been shown to play an important role in many cancers [7–11], but its levels of expression, clinical, pathological characteristics and survival in patients with CRC still understudied.

Although gene expression profiling has revealed several molecular changes associated with CRC [32,33]. Biological markers that precisely predict progression and prognosis are not yet available.

In the current study, mir-221 was over expressed significantly in CRC samples compared with adjacent non-cancerous tissue samples; this increase was accompanied by advanced clinical stage, large tumor size with local infiltration, lymph node metastasis and poor survival.

Our results are in agreement with other studies. Liu et al. demonstrated that human CRC tissues had higher levels of mir-221 than non-tumor colon tissues [34]. Yau et al. also reported that miR-221 levels were significantly higher in 40 CRC tumors compared with their respective adjacent normal tissues and miR-221 showed a significant rise in the trend from normal control to

Table 1. Clinical and pathological features in colorectal cancer patients and mir-221 expression levels

Clinical and pathological parameters	No.	miR-221 (ΔC_T) mean \pm SD	P
Age			
< 60 years	48	2.38 \pm 0.16	NS*
\geq 60 years	44	2.70 \pm 0.25	
Gender			
Male	49	2.54 \pm 0.33	NS*
Female	43	2.42 \pm 0.16	
Differentiation grade			
G ₁ -G ₂	40	2.28 \pm 0.24	NS*
G ₃ -G ₄	52	2.89 \pm 0.23	
Stage			
I-II	56	2.18 \pm 0.14	P = 0.018
III	36	3.09 \pm 0.32	
T			
T ₁ -T ₂	62	2.09 \pm 0.12	P = 0.006
T ₃	30	3.17 \pm 0.46	
LNs			
Negative	30	2.14 \pm 0.16	P = 0.032
Positive	62	2.92 \pm 0.34	

*NS, not statistically significant (P \geq 0.05, Student's t-test)**Table 2. Univariate Cox proportional hazard regression analyses of the relationship between clinical-pathological characteristics and levels of mir-221**

Variable		Hazard ratio (HR)	P
Age	\geq 60 vs < 60 years	0.678	NS
Gender	Male vs female	0.435	0.23
Level of mir-221 expression	High vs low	2.422	0.016
Tumor size and infiltration (T)	T1+T2 vs T3	5.652	< 0.001
Tumor stage (TNM)	I +II vs III	3.462	0.008
Lymph node metastasis	+ve vs -ve	2.956	0.006
Degree of tumor differentiation	G1 + GII vs GIII + GIV	5.165	0.011

Table 3. Multivariate Cox proportional hazard regression analyses of the relationship between clinical-pathological characteristics and levels of mir-221

Variable		Hazard ratio (HR)	P
Age	\geq 60 vs < 60 years	0.725	NS
Gender	Male vs female	0.571	NS
Level of mir-221 expression	High vs low	2.321	0.024
Tumor size and infiltration (T)	T1+T2 vs T3	4.268	0.001
Tumor stage (TNM)	I +II vs III	3.225	0.016
Lymph node metastasis	+ve vs -ve		0.006
Degree of tumor differentiation	G1 + GII vs GIII + GIV		0.011

late CRC (P<0.0001). The levels of stool mir-221 are significantly higher in subjects with stage I+II and stage III+IV CRC compared to normal controls [35]. miR-221 down regulated in CRC and its level was higher in patients with liver metastases in comparison with

nonmetastatic patients [36]. In addition Liu et al. show the predictive value of mir-221 in the prognosis of colorectal cancer (CRC) and its effects on CRC cell proliferation, migration, and invasion. Mir-221 expression was associated with tumor size, TNM stage, Duke stage, LNM,

local recurrence rate, and distant metastases [37]. Liao et al. studied survival using Cancer Genome Atlas data indicated that increased expression of mir-221 was associated with lower survival in CRC patients [38]. Tao et al. [39] found that high levels of mir-221 is associated with a more tumor phenotype and decreased time to recur after surgery in CRC patients. Gramantieri *et al.* also found the same previous results but in patients with hepatocellular carcinoma [22]. Sun et al. noticed that up-regulated mir-221 expression significantly with deep local invasion and advanced TNM stage in CRC [32].

Molecular mechanisms that link mir-221 over expression to cancer and their functional targets have been identified and revealed by several studies. It is known that the progression of cancer cells from G0/G1 to S phase in the cell cycle is promoted by mir-221, it also facilitates cell proliferation by down-regulates p27Kip1 and p57Kip2. Both are tumor suppressors and cell-cycle inhibitors related to the protein family kinase inhibitors [13,40,41].

It is also has been proofed the common oncogenic protein, c-Kit, is down streaming mir-221 in gastrointestinal stromal tumors (GIST) [42], papillary thyroid carcinoma [43], prostate cancer [44] and leukemic cells [45].

One miR may have more than 100 targets [46] and a single mRNA may be regulated by a number of miRs, this means that mir-221 induced mechanisms of tumor genesis and development are not understood [47].

5. CONCLUSION

In conclusion, the present study confirmed the over expression of mir-221 in CRC tissue samples in comparison with normal adjacent non- cancerous tissue samples. It also showed a relation between the up-regulation of mir-221 and tumor infiltration, progression and poor prognosis in CRC. Our findings suggest mir-221 may be used as a prognostic marker in CRC and it needs more studies to be used as a potential diagnostic marker and as a molecular target for targeted cancer therapy.

CONSENT

Informed consent was obtained from all individual participants included in the study.

ETHICAL APPROVAL

This study was carried out in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards. This study was approved by the ethical committee of Zagazig University.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Hagggar FA, Boushey RP. Colorectal cancer epidemiology: Incidence, mortality, survival, and risk factors. *Clin. Colon Rectal Surg.* 2009;22:191–197. DOI: 10.1055/s-0029-1242458
2. Siegel R, Desantis C, Jemal A. Colorectal cancer statistics. *CA Cancer J. Clin.* 2014;64:104–117. DOI: 10.3322/caac.21220
3. Edwards BK, Ward E, Kohler BA, Ehemann C, Zauber AG, Anderson RN et al. Annual report to the nation on the status of cancer, 1975–2006, featuring colorectal cancer trends and impact of interventions (risk factors, screening, and treatment) to reduce future rates. *Cancer.* 2010;116: 544–573. DOI: 10.1002/cncr.24760
4. Howlader N, Noone AM, Krapcho M, Garshell J, Miller D, Altekruse SF et al. *SEER Cancer Statistics Review, 1975–2012.* National Cancer Institute; Bethesda, MD, USA: 2015. [Accessed on 30 July 2015] Available: http://seer.cancer.gov/csr/1975_2012/
5. Rupaimoole R, Slack FJ. MicroRNA therapeutics: Towards a new era for the management of cancer and other diseases. *Nat Rev Drug Discov.* 2017;16: 203–222. DOI: 10.1038/nrd.2016.246
6. Silvia Galardi, Neri Mercatelli, Ezio Giorda, Simone Massalini, Giovanni Vanni Frajese, Silvia Anna Ciafrè et al. miR-221 and miR-222 Expression Affects the Proliferation Potential of Human Prostate Carcinoma

- Cell Lines by Targeting p27Kip1. *J. Biol. Chem.* 2007;282:23716.
DOI: 10.1074/jbc.M701805200
7. Peadar S. Waters, Ailbhe M. McDermott, Deirdre Wall, Helen M. Heneghan, Nicola Miller, John Newell et al. Relationship between circulating and tissue microRNAs in a Murine model of breast cancer. *Plos One.* 2012;7(11):e50459.
Available:<https://doi.org/10.1371/journal.pone.0050459>
 8. Nassirpour R, Mehta PP, Baxi SM, Yin MJ. miR-221 Promotes Tumorigenesis in Human Triple Negative Breast Cancer Cells. *Plos One.* 2013;8(4):e62170.
 9. Zhihong Wang, Hao Zhang, Liang He, Wenwu Dong, Jing Li et al. Association between the expression of four upregulated miRNAs and extrathyroidal invasion in papillary thyroid carcinoma. *Onco Targets Ther.* 2013;6:281–287.
 10. Yang J, Zhang J-y, Chen J, Xu Y, Song N-h, Yin C-j. Prognostic Role of MicroRNA-221 in Various Human Malignant Neoplasms: A Meta-Analysis of 20 Related Studies. *Plos One.* 2014;9(1):e87606.
Available:<https://doi.org/10.1371/journal.pone.0087606>
 11. Sun, Tong, Ming Yang, Philip W. Kantoff, Gwo-Shu Mary Lee. Role of MicroRNA-221/-222 in Cancer Development and Progression. *Cell Cycle.* 2009;8(15):2315–16.
Available:<https://doi.org/10.4161/cc.8.15.9221>
 12. Kawaguchi T, Komatsu S, Ichikawa D, Morimura R, Tsujiura M, Konishi H, et al. Clinical impact of circulating miR-221 in plasma of patients with pancreatic cancer. *Br J Cancer* 2013;108(2):361.
 13. Galardi S, Mercatelli N, Farace MG, Ciafre SA. NF-kB, c-Jun induce the expression of the oncogenic miR-221 and miR-222 in prostate carcinoma and glioblastoma cells. *Nucleic Acids Res.* 2011;39(9):3892-902.
 14. Hong F, Li Y, Xu Y, Zhu L. Prognostic significance of serum microRNA-221 expression in human epithelial ovarian cancer. *J Int Med Res.* 2013;41(1):64-71.
 15. Papaconstantinou IG, Manta A, Gazouli M, Lyberopoulou A, Lykoudis PM, Polymeneas G, et al. Expression of microRNAs in patients with pancreatic cancer and its prognostic significance. *Pancreas.* 2013;42(1):67-71.
 16. Nassirpour R, Mehta PP, Baxi SM, Yin MJ. miR-221 promotes tumorigenesis in human triple negative breast cancer cells. *PloS one.* 2013;8(4):e62170.
 17. Wang Z, Zhang H, He L, Dong W, Li J, Shan Z et al. Association between the expression of four upregulated miRNAs and extrathyroidal invasion in papillary thyroid carcinoma. *Onco Targets Ther.* 2013;6:281.
 18. Di Martino MT, Gullà A, Cantafio ME, Lionetti M, Leone E, Amodio N, et al. In vitro and in vivo anti-tumor activity of miR-221/222 inhibitors in multiple myeloma. *Oncotarget.* 2013;4(2):242.
 19. Pineau P, Volinia S, McJunkin K, Marchio A, Battiston C, Terris B, et al. miR-221 over expression contributes to liver tumorigenesis. *Proc Natl Acad Sci.* 2010; 107(1):264-9.
 20. Zhang C, Zhang J, Hao J, Shi Z, Wang Y, Han L, et al. High level of miR-221/222 confers increased cell invasion and poor prognosis in glioma. *J Transl Med.* 2012; 10(1):119.
 21. Liu K, Li G, Fan C, Diao Y, Wu B, Li J. Increased Expression of MicroRNA-221 in gastric cancer and its clinical significance. *J Int Med Res.* 2012;40(2): 467-74.
 22. Gramantieri L, Fornari F, Ferracin M, Veronese A, Sabbioni S, Calin GA, et al. MicroRNA-221 targets Bmf in hepatocellular carcinoma and correlates with tumor multifocality. *Clin Cancer Res.* 2009;15(16):5073-81.
 23. Li J, Wang Y, Yu W, Chen J, Luo J. Expression of serum miR-221 in human hepatocellular carcinoma and its prognostic significance. *Biochem Biophys Res Commun.* 2011;406(1):70-3.
 24. Zhou Y-L, Liu C, Dai X, Zhang X-H, Wang O-C. Over expression of miR-221 is associated with aggressive clinicopathologic characteristics and the BRAF mutation in papillary thyroid carcinomas. *Medical Oncology.* 2012;29(5):3360–6.
 25. Yoon SO, Chun SM, Han EH, Choi J, Jang SJ, Koh SA, et al. Deregulated expression of microRNA-221 with the potential for prognostic biomarkers in surgically resected hepato cellular carcinoma.

- Human Pathology. 2011;42(10):1391-400.
26. Kang SG, Ha YR, Kim SJ, Kang SH, Park HS, Lee JG, et al. Do microRNA 96, 145 and 221 expressions really aid in the prognosis of prostate carcinoma? *Asian J Androl.* 2012;14(5):752.
 27. Cardinali B, Castellani L, Fasanaro P, Basso A, Alema S, Martelli F, et al. MicroRNA-221 and microRNA-222 modulate differentiation and maturation of skeletal muscle cells. *PLoS One.* 2009;4(10):e7607.
 28. Amin MB, Greene FL, Edge SB, Compton CC, Gershenwald JE, Brookland RK, Meyer L, Gress DM, Byrd DR, Winchester DP. The Eighth Edition AJCC Cancer Staging Manual: Continuing to build a bridge from a population-based to a more personalized approach to cancer staging. *CA: A Cancer Journal for Clinicians.* 2017; 67(2):93-9.
 29. Lawrie CH, Gal S, Dunlop HM, Pushkaran B, Liggins AP, Pulford K, et al. Detection of elevated levels of tumor-associated microRNAs in serum of patients with diffuse large B-cell lymphoma. *British Journal of Haematology.* 2008;141(5):672-5.
 30. Hegde SR, Sun W, Lynch JP. Systemic and targeted therapy for advanced colon cancer. *Expert Rev Gastroenterol Hepatol.* 2008;2:135-149.
 31. Kozomara A, Griffiths-Jones S. miRBase: annotating high confidence microRNAs using deep sequencing data. *Nucleic Acids Res.* 2014;42:68-73.
 32. Sun K, Wang W, Zeng JJ, Wu CT, Lei ST, Li GX. MicroRNA-221 inhibits CDKN1C/p57 expression in human colorectal carcinoma. *Acta Pharmacologica Sinica.* 2011;32(3):375.
 33. Qin J, Luo M. MicroRNA-221 promotes colorectal cancer cell invasion and metastasis by targeting RECK. *FEBS Lett.* 2014;588:99-104.
 34. Liu S, Sun X, Wang M, Hou Y, Zhan Y, Jiang Y, Liu Z, Cao X, Chen P, Liu Z, Chen X. A microRNA 221-and 222-mediated feedback loop maintains constitutive activation of NFκB and STAT3 in colorectal Cancer cells. *Gastroenterology.* 2014; 147(4):847-59.
 35. Yau TO, Wu CW, Dong Y, Tang CM, Ng SS, Chan FK, Sung JJ, Yu J. microRNA-221 and microRNA-18a identification in stool as potential biomarkers for the non-invasive diagnosis of colorectal carcinoma. *British Journal of Cancer.* 2014;111(9): 1765.
 36. Hur K, Toiyama Y, Schetter AJ, Okugawa Y, Harris CC, Boland CR, Goel A. Identification of a metastasis-specific MicroRNA signature in human colorectal cancer. *Journal of the National Cancer Institute.* 2015;107(3):dju492.
 37. Liu L, Meng T, Yang XH, Sayim P, Lei C, Jin B, Ge L, Wang HJ. Prognostic and predictive value of long non-coding RNA GAS5 and microRNA-221 in colorectal cancer and their effects on colorectal cancer cell proliferation, migration, and invasion. *Cancer Biomarkers.* 2018;22(2): 283-99.
 38. Liao D, Li T, Ye C, Zeng L, Li H, Pu X, Ding C, He Z, Huang GL. miR 221 inhibits autophagy and targets TP53INP1 in colorectal cancer cells. *Experimental and Therapeutic Medicine.* 2018;15(2):1712-7.
 39. Tao K, Yang J, Guo Z, Hu Y, Sheng H, Gao H, et al. Prognostic value of miR-221-3p, miR-342-3p and miR-491-5p expression in colon cancer. *Am J Transl Res.* 2014;6(4):391.
 40. Medina R, Zaidi SK, Liu CG, Stein JL, Croce CM, Stein GS. MicroRNAs 221 and 222 bypass quiescence and compromise cell survival. *Cancer Res.* 2008;68(8): 2773-80.
 41. F Fornari, L Gramantieri, M Ferracin, A Veronese, S Sabbioni, G A Calin, et al. MiR-221 controls CDKN1C/p57 and CDKN1B/p27 expression in human hepatocellular carcinoma. *Oncogene.* 2008;27:5651.
 42. Koelz M, Lense J, Wrba F, Scheffler M, Dienes HP, Odenthal M. Down-regulation of miR-221 and miR-222 correlates with pronounced Kit expression in gastrointestinal stromal tumors. *Int J Oncol.* 2011;38(2):503-11.
 43. He H, Jazdzewski K, Li W, Liyanarachchi S, Nagy R, Volinia S, et al. The role of microRNA genes in papillary thyroid carcinoma. *Proceedings of the National Academy of Sciences.* 2005;102(52): 19075-80.
 44. Spahn, M, Kneitz S, Scholz CJ, Stenger N, Rüdiger T, Ströbel P et al. Expression of

- microRNA-221 is progressively reduced in aggressive prostate cancer and metastasis and predicts clinical recurrence. *Int J Cancer*. 2010;127(2):394-403.
DOI: 10.1002/ijc.24715
45. Felli N, Fontana L, Pelosi E, Botta R, Bonci D, Facchiano F et al. MicroRNAs 221 and 222 inhibit normal erythropoiesis and erythroleukemic cell growth via kit receptor down-modulation. *Proc Natl Acad Sci*. 2005;102(50):18081-6.
46. Brennecke J, Stark A, Russell RB, Cohen SM. Principles of microRNA–target recognition. *PLoS Biol*. 2005;3(3):e85.
47. Johnson SM, Grosshans H, Shingara J, Byrom M, Jarvis R, Cheng A, et al. RAS is regulated by the let-7 microRNA family. *Cell*. 2005;120(5):635-47.

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